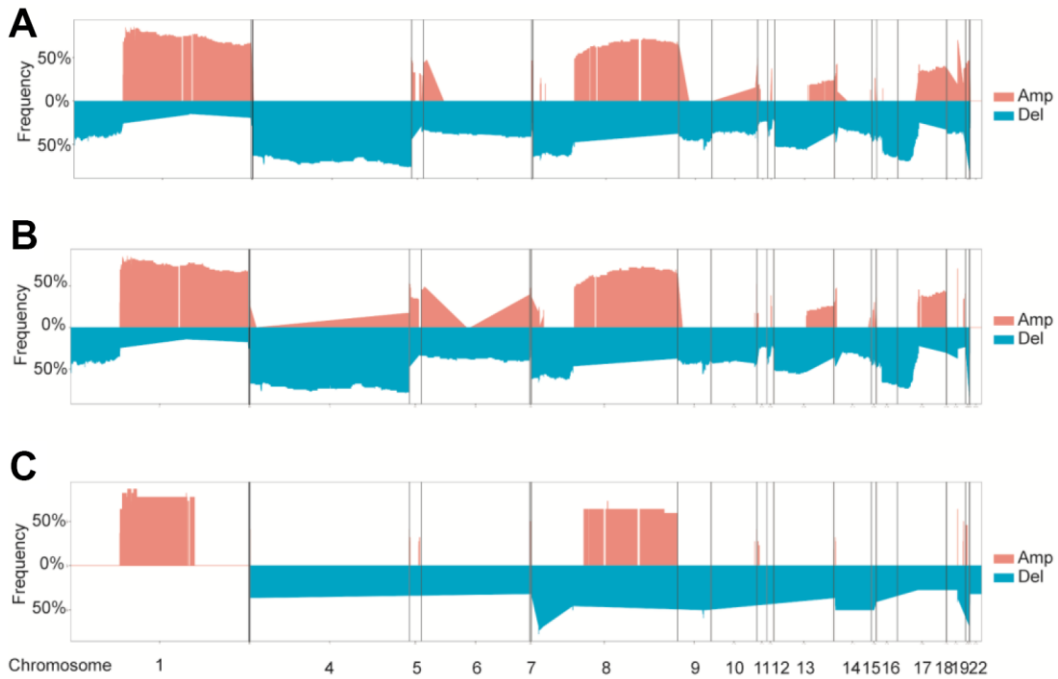
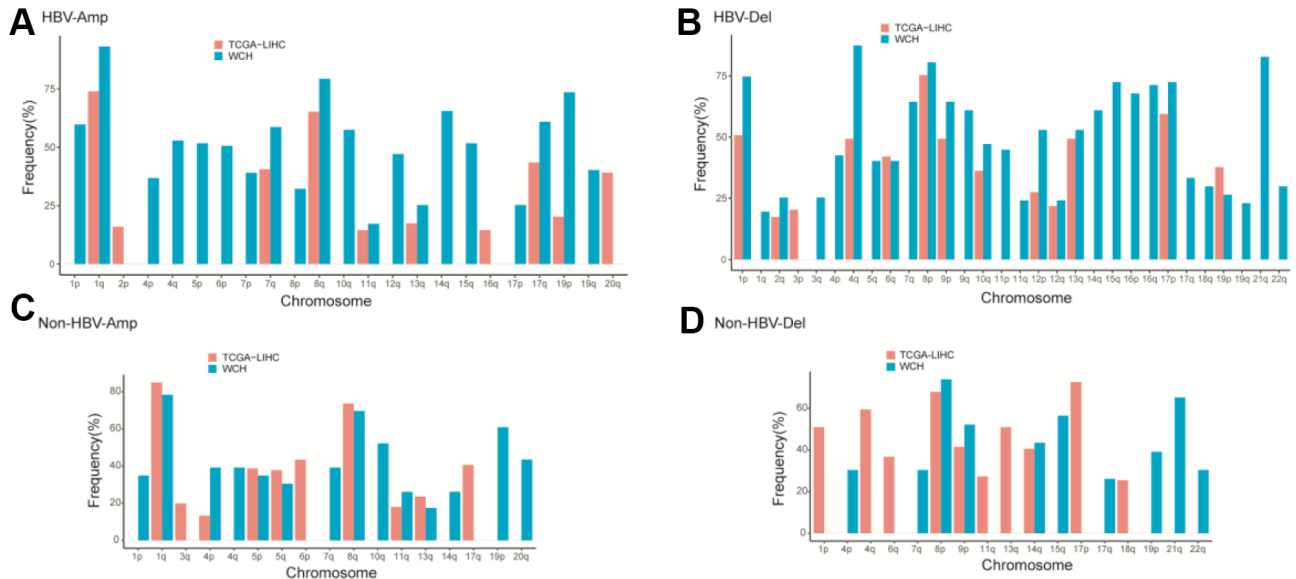


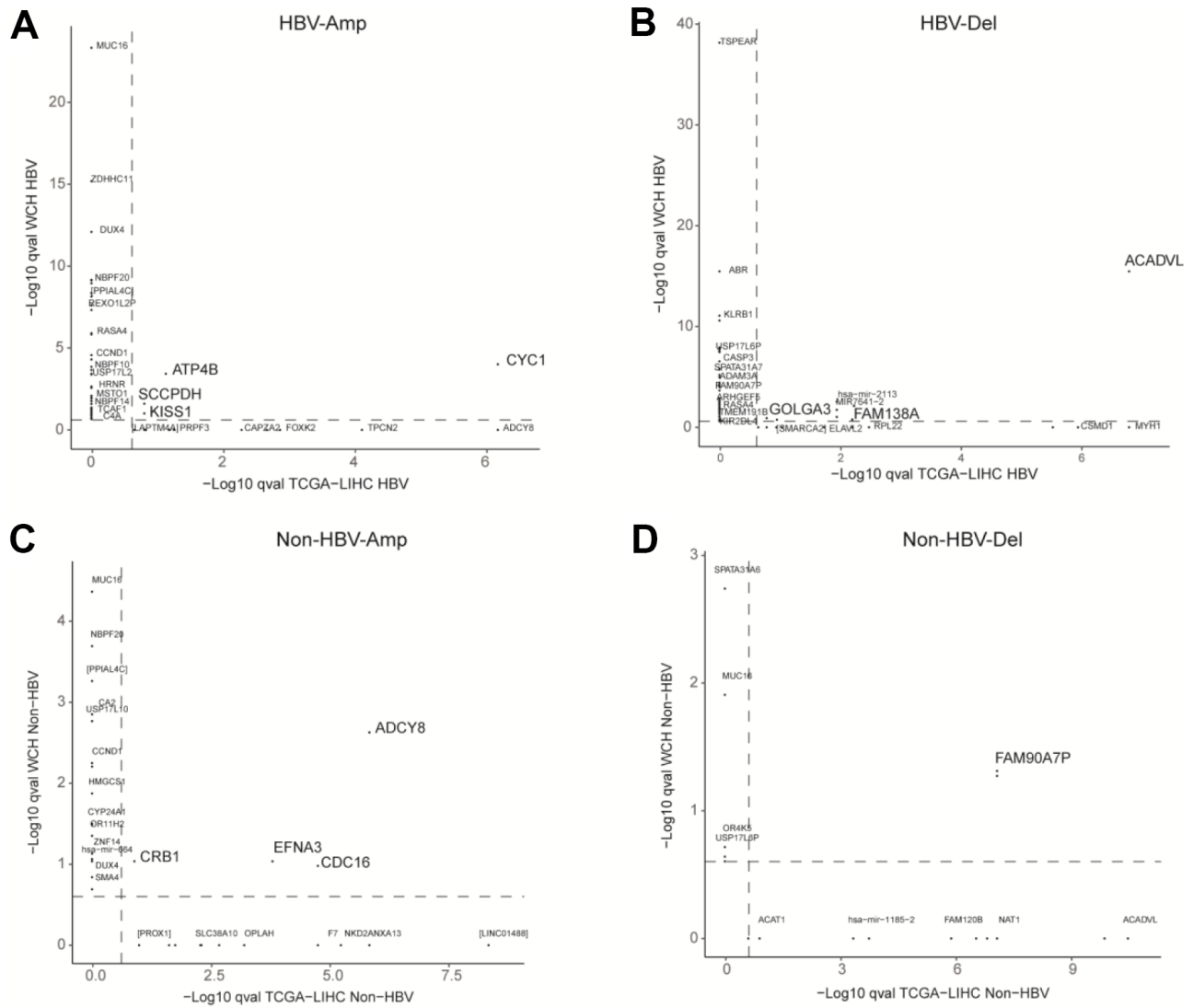
**SUPPLEMENTARY FIGURES**



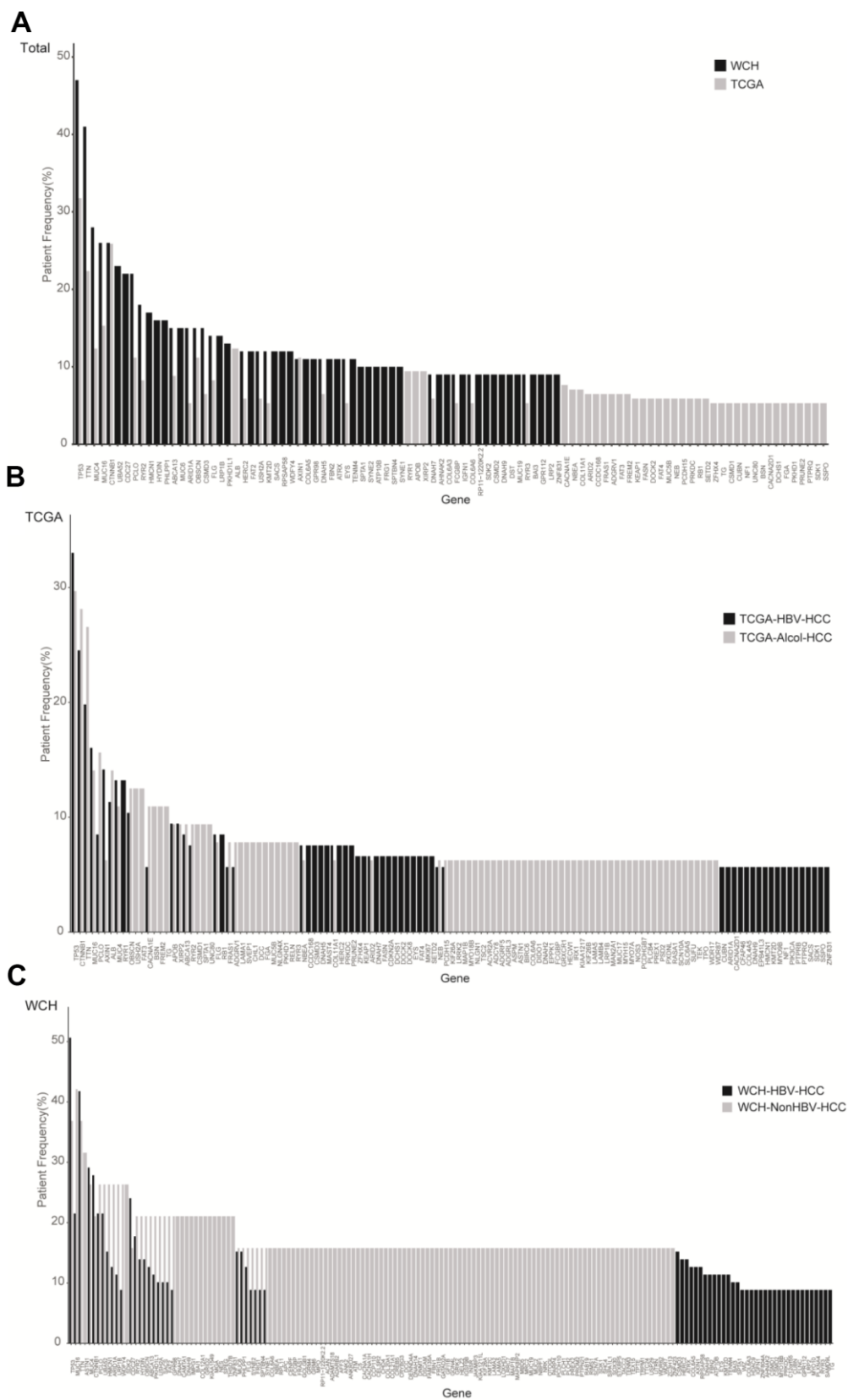
**Supplementary Figure 1.** Overview of copy number variation in the WCH group (A) All patients in the WCH group; (B) Patients with hepatitis B (WCH-HBV-HCC group); (C) Patients without hepatitis B (WCH-NonHBV-HCC group). The red color represents copy number amplification, and the green represents copy number deletion.



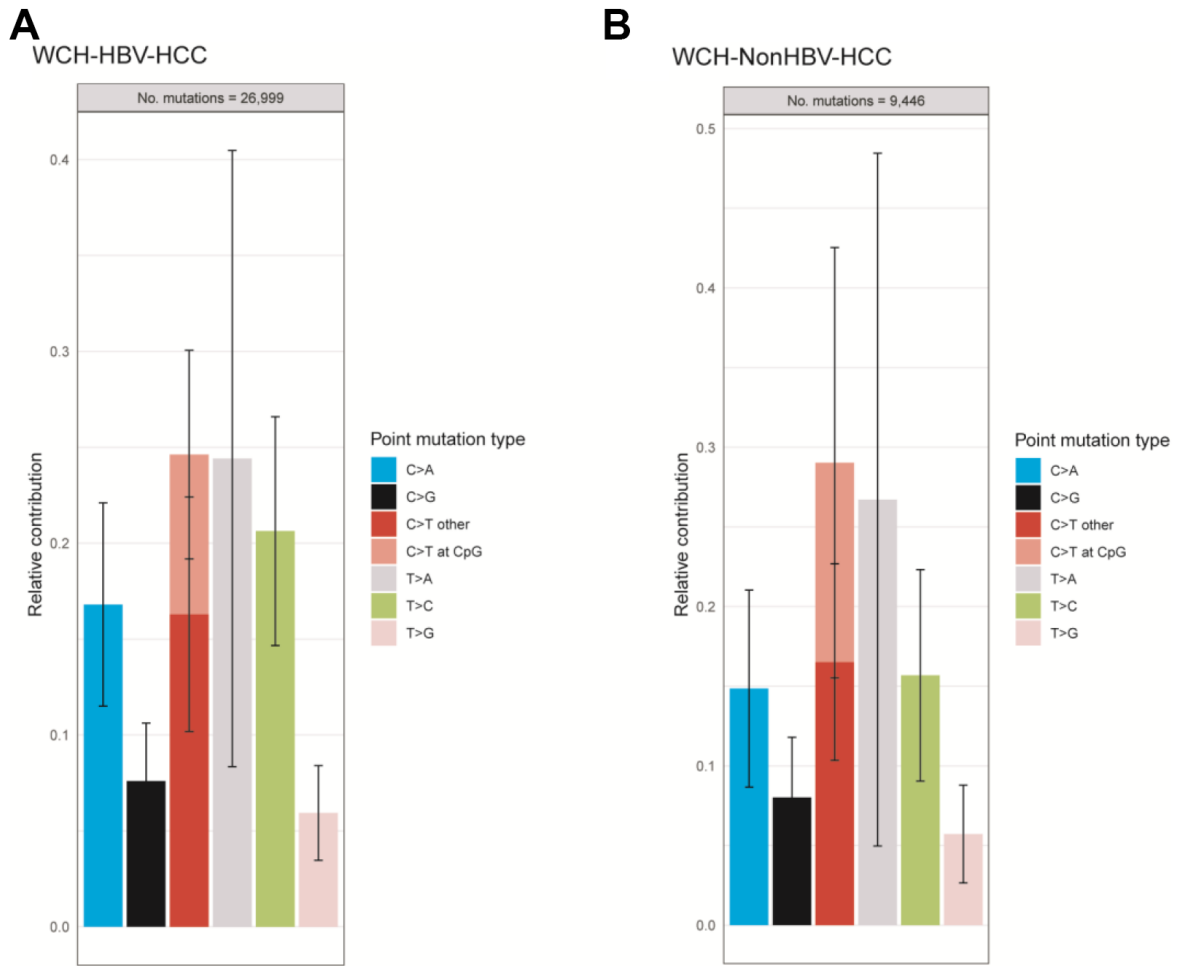
**Supplementary Figure 2. Comparison of arm level copy number alterations between WCH and TCGA cohorts.** (A) Amplification frequencies of WCH-HBV-HCC versus TCGA-HBV-HCC group; (B) Deletion frequencies of WCH-HBV-HCC versus TCGA-HBV-HCC group; (C) Amplification frequencies of WCH-NonHBV-HCC versus TCGA-Alcol-HCC group; (D) Deletion frequencies of TCGA-Alcol-HCC versus WCH-NonHBV-HCC group.



**Supplementary Figure 3. The focal CNV profile between the WCH and TCGA were compared to identify novel focal events.** (A) Focal amplifications in the WCH-HBV-HCC and TCGA-HBV-HCC groups. (B) Focal deletions in the WCH-HBV-HCC and TCGA-HBV-HCC groups. (C) Focal amplifications in the WCH-NonHBV-HCC and TCGA-NonHBV-HCC groups. (D) Focal deletions in the WCH-NonHBV-HCC and TCGA-NonHBV-HCC groups. The q values for amplifications (A, C) and deletions (B, D) in the WCH group were plotted against q values from the TCGA cohort. CNVs with q values <0.25 were deemed as significant. Owing to the similar q values of a large number of genes, we only showed parts of representative genes in this figure. All shared and unique genes among the above groups are shown in the Supplementary data file 2-5. The dashed line are q value cutoffs.

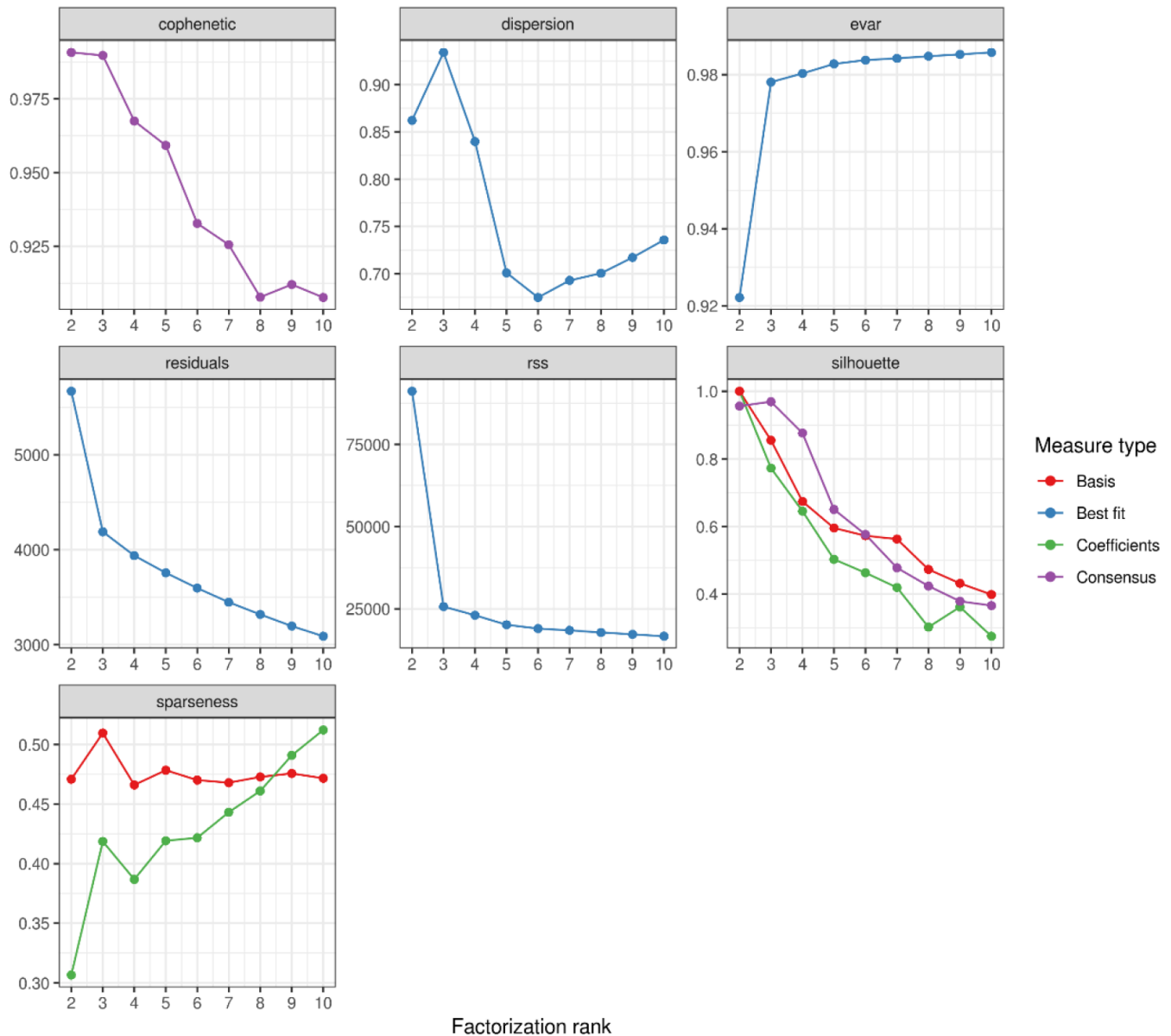


**Supplementary Figure 4. Comparison of the mutation frequencies of significantly mutated gene between the WCH and TCGA cohorts. (A) The total WCH and TCGA cohorts; (B) TCGA-HBV-HCC and TCGA-Alcoi-HCC groups; (C) WCH-HBV-HCC and WCH-NonHBV-HCC groups.**



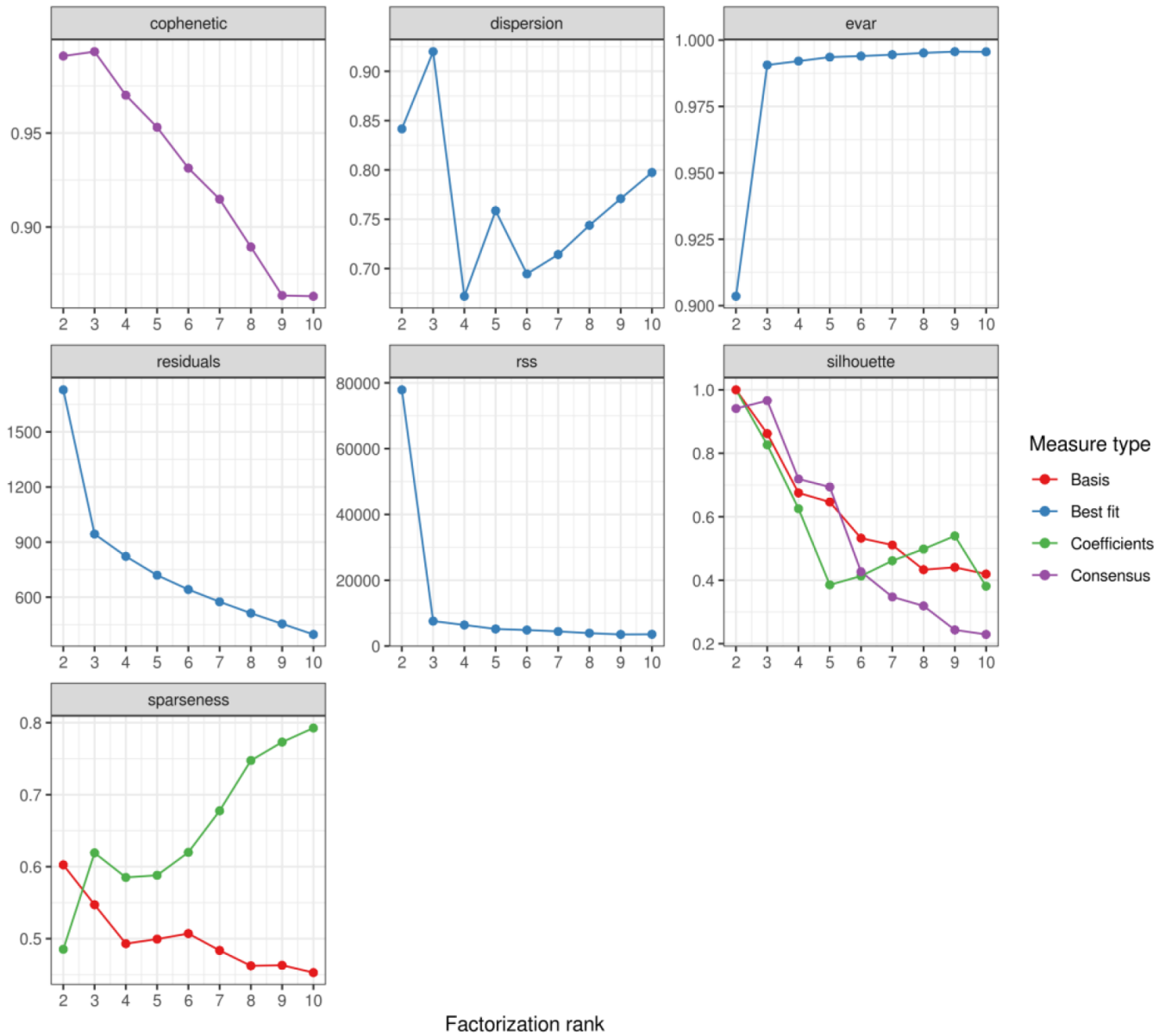
**Supplementary Figure 5. Mutation spectrum of patients in WCH group. (A) WCH-HBV-HCC group; (B) WCH-NonHBV-HCC group.**

## NMF rank survey

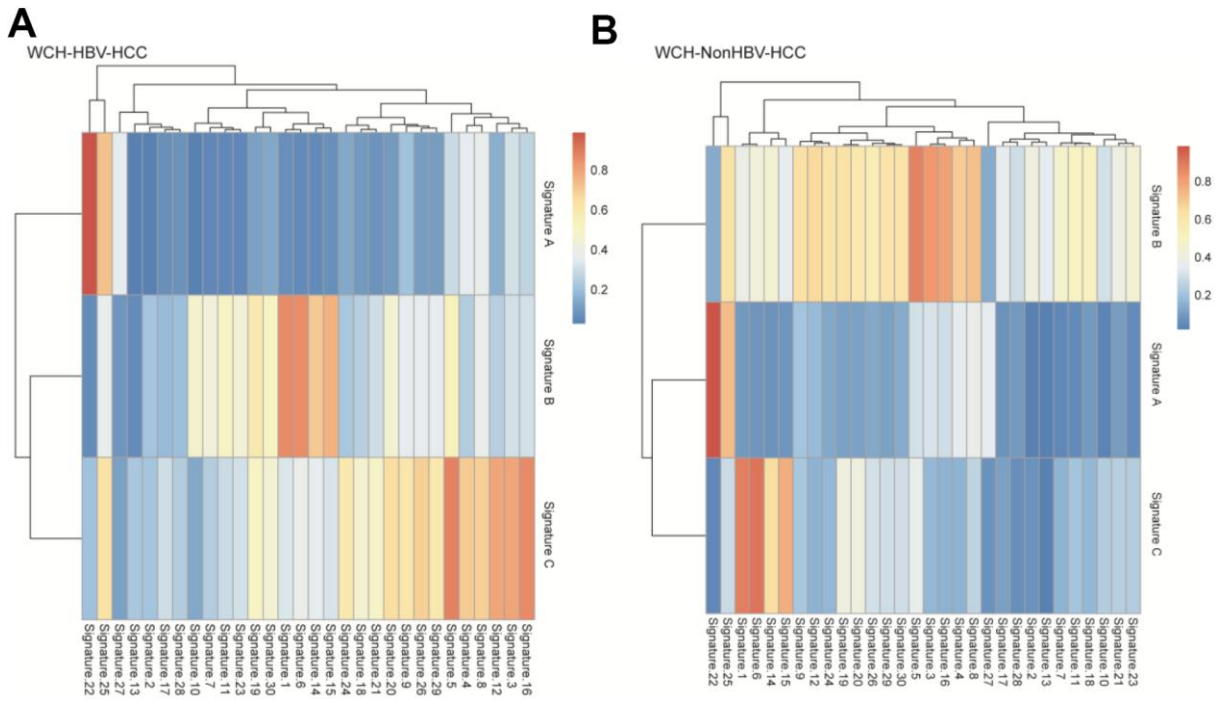


**Supplementary Figure 6. Calculating the optimal clustering value based on the NMF algorithm in the WCH-HBV-HCC group.** Cophenetic refers to correlation coefficient; Dispersion is the dispersion coefficient (evaluation of the repeatability of the NMF results); Evar is used to evaluation of the interpretation effect of the NMF model to the matrix differences; Silhouette is aimed to evaluate the stability of the model; Sparseness is used to calculate the sparsity of the matrix. RSS, Residual Sum of Squares.

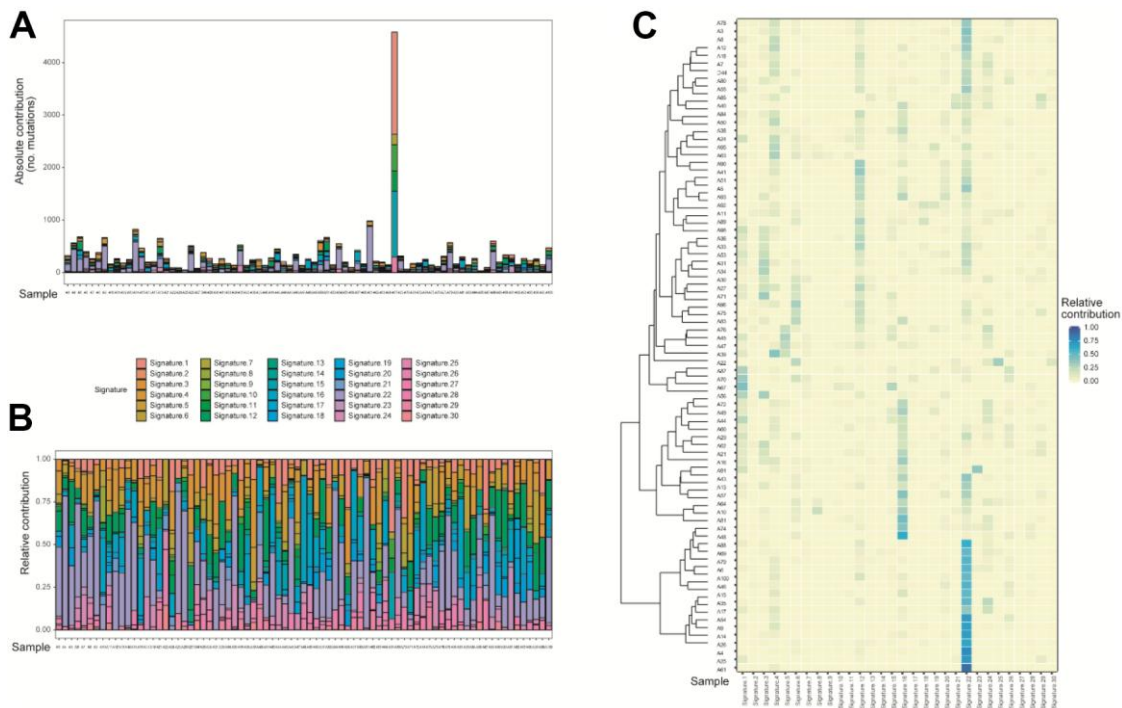
### NMF rank survey



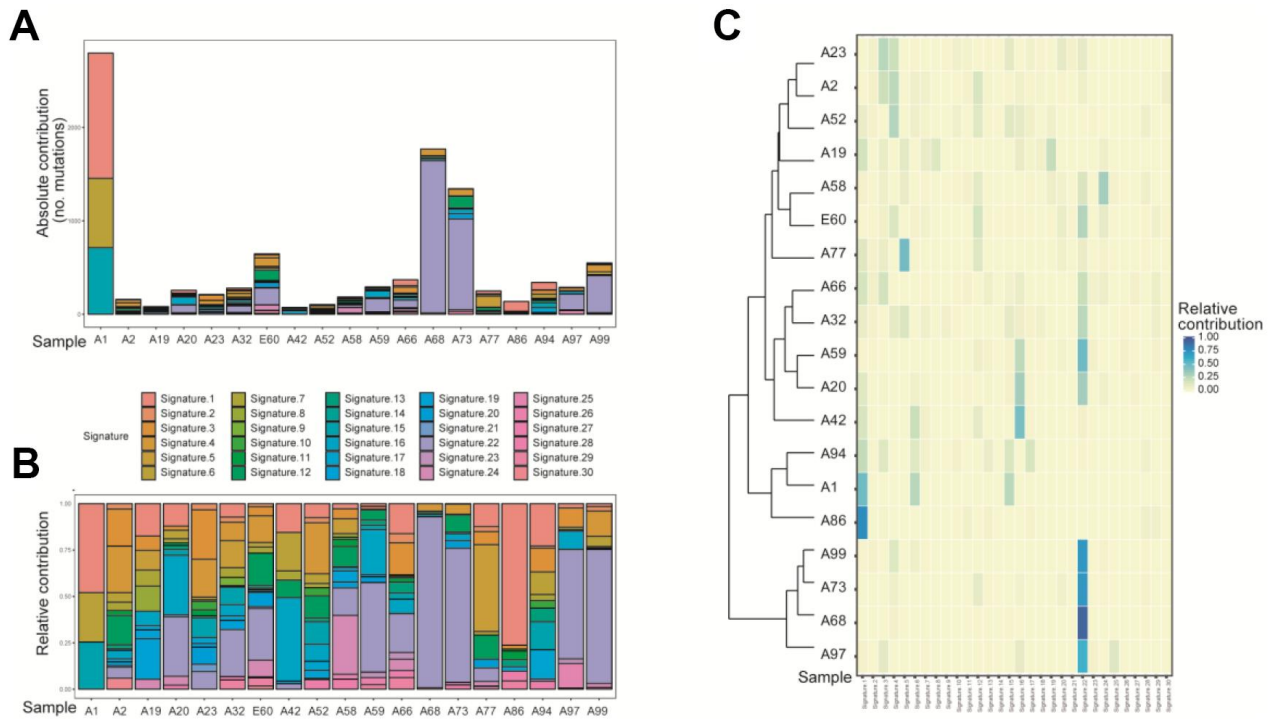
**Supplementary Figure 7. Calculating the optimal clustering value based on the NMF algorithm in the WCH-NonHBV-HCC group.** Cophenetic refers to correlation coefficient; Dispersion is the dispersion coefficient (evaluation of the repeatability of the NMF results); Evar is used to evaluation of the interpretation effect of the NMF model to the matrix differences; Silhouette is aimed to evaluate the stability of the model; Sparseness is used to calculate the sparsity of the matrix. RSS, Residual Sum of Squares.



**Supplementary Figure 8. Association of the mutation Signatures identified in this study with the existing mutation signatures of the COSMIC database. (A) WCH-HBV-HCC group; (B) WCH-NonHBV-HCC group.**

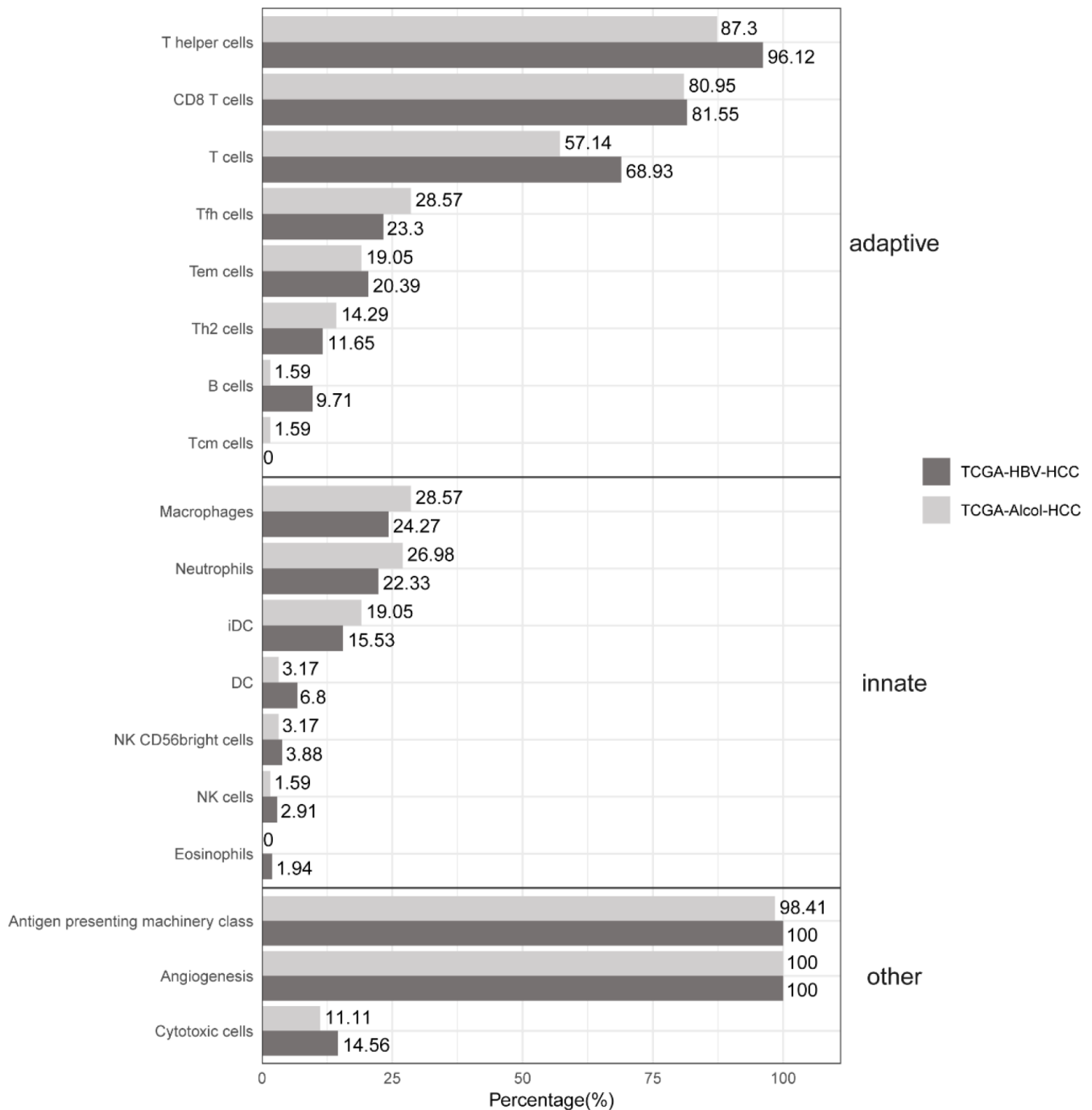


**Supplementary Figure 9. Distribution of the 30 mutation Signatures of the COSMIC database among all samples in the WCH-HBV-HCC group (A) The contributions of the 30 mutational signatures to tumors in the WCH-HBV-HCC group. The sample names are displayed on the horizontal axis, whereas the vertical axis depicts the number of mutations of samples in the WCH-HBV-HCC group; (B) The relative contribution of the 30 Signatures in samples from the WCH-HBV-HCC group; (C) The distribution of the 30 mutation Signatures in the WCH-HBV-HCC group.**

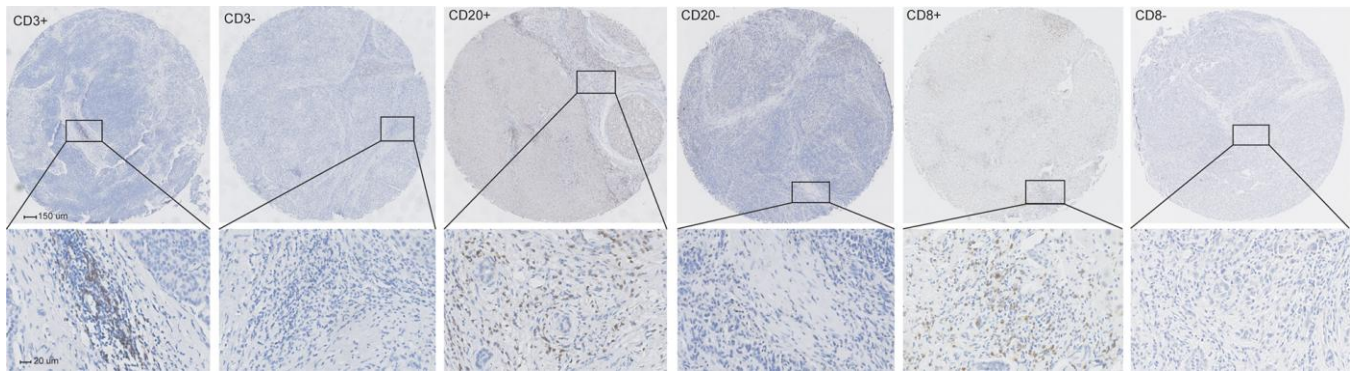


**Supplementary Figure 10.** Distribution of the 30 mutation Signatures of the COSMIC database among all samples in the WCH-NonHBV-HCC group (**A**) The contributions of the 30 mutational signatures to tumors in the WCH-NonHBV-HCC group. The sample names are displayed on the horizontal axis, whereas the vertical axis depicts the number of mutations of samples in the WCH-NonHBV-HCC group; (**B**) The relative contribution of the 30 Signatures in samples from the WCH-NonHBV-HCC group; (**C**) The distribution of the 30 mutation Signatures in the WCH-NonHBV-HCC group.

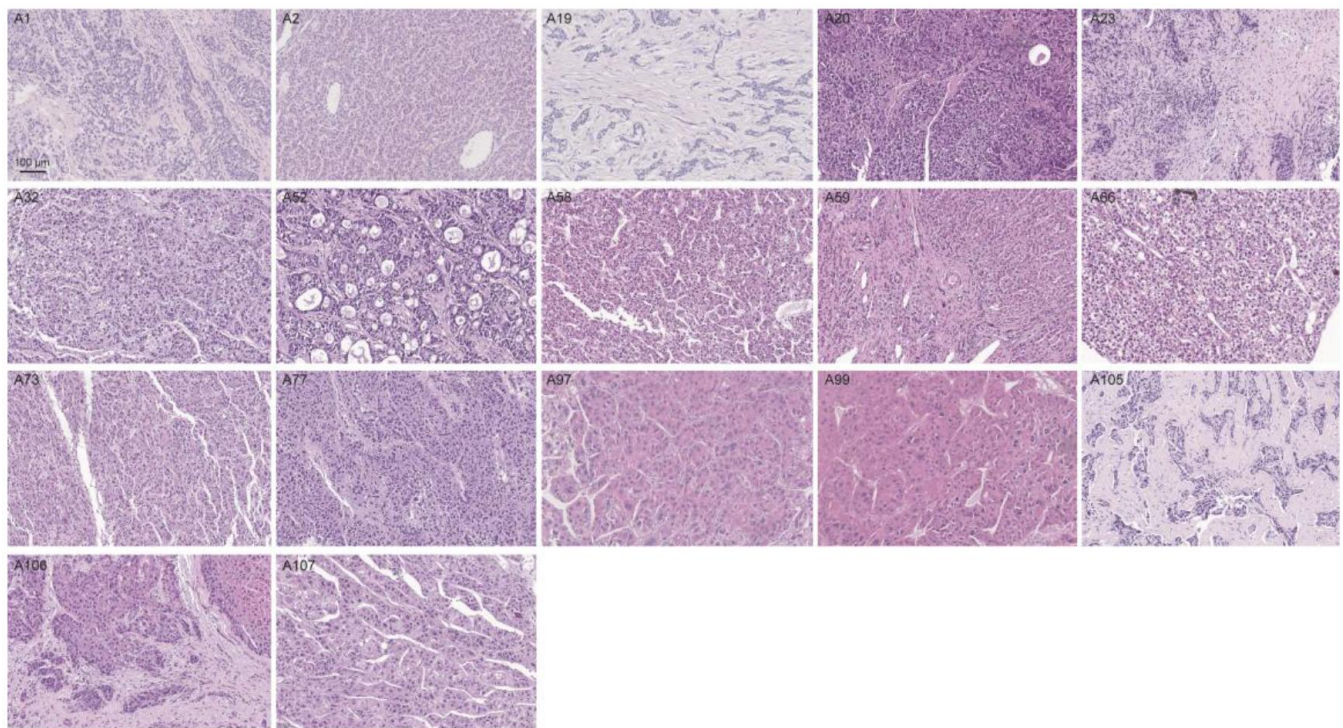




**Supplementary Figure 11. Comparison of Tumor-infiltrating lymphocytes enrichment profile in the TCGA-HBV-HCC group and TCGA-Alcol-HCC group.** Percentage of cases with enriched immune cell signatures were calculated using the GSVA and pre-rank Gene Set Enrichment Analysis (GSEA) methods (see Supplementary Methods). The GSEA was utilized to calculate enrichment score, while the pre-rank GSEA was used to calculate FDR values and for each immune cell signature, enrichment is defined as  $q$ -value  $\leq 0.1$ . The black bars indicate the percentage of patients having significant enrichment for the given immune cell type in the TCGA-HBV-HCC group, while gray bars represent the percentage in the TCGA-Alcol-HCC group. Immune cell signatures were classified into adaptive, innate and other. Source data are provided in the Supplementary data file 16.



**Supplementary Figure 12. Representative images of immunohistochemical staining of HCC samples in the WCH group.** These immune markers are CD3+ T cells, CD20+ T cells and CD8+ B cells.



**Supplementary Figure 13. Representative images of hematein-eosin staining of HCC samples in the WCH-NonHBV-HCC group.** The pathological features for samples in the WCH-NonHBV-HCC is distinct, and these different pathological patterns include pseudoglandular histological pattern (A52), fibrolamellar-HCC (A19), trabecular histological pattern (A107) and steatosis (A66), etc.