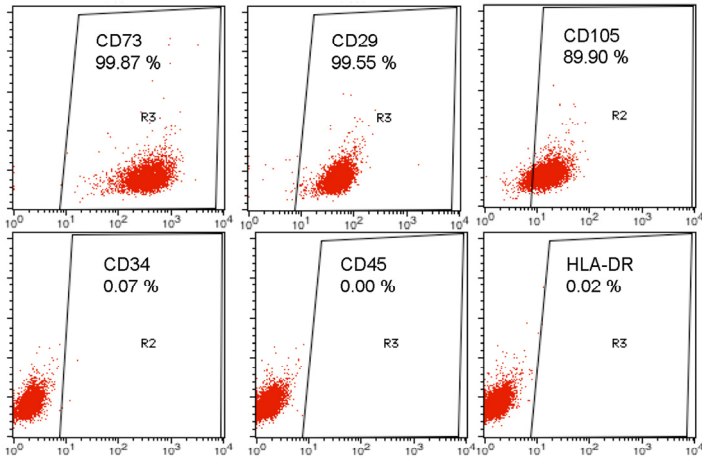
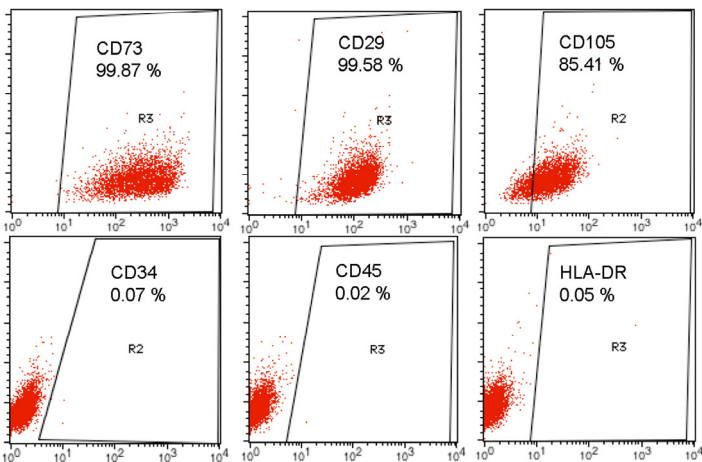


SUPPLEMENTARY DATA

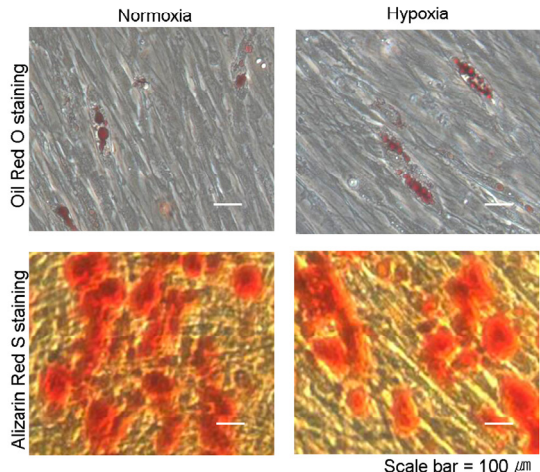
A



B



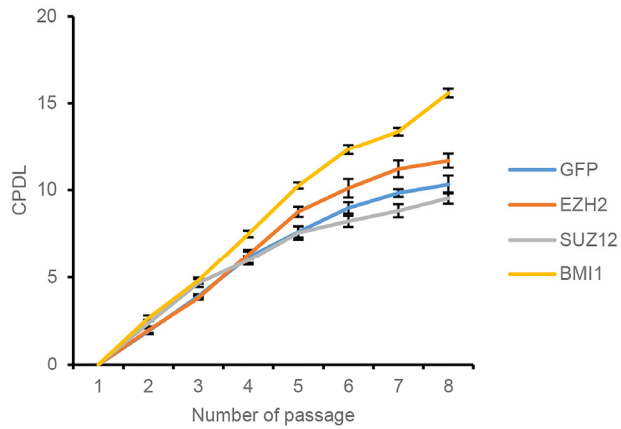
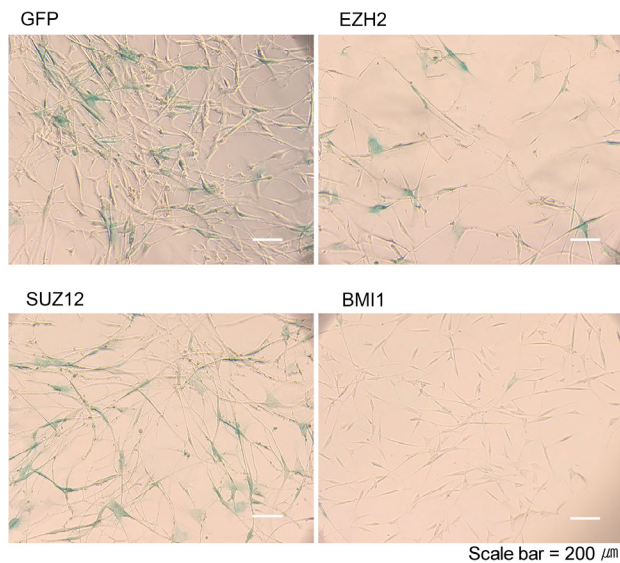
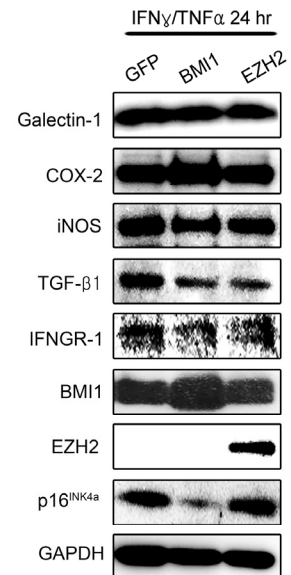
C



Supplementary Figure S1. Characterization of hUCB-MSCs cultured in normoxia and hypoxia. Normoxic (A)- and hypoxic (B)-cultured hUCB-MSCs (1×10^6 cells/ml) were stained with FITC- or PE-conjugated antibodies specific for human CD29, CD34, CD45, CD73, CD105, and HLA-DR. (C) Images of differentiated hUCB-MSCs after induction into specific tissues. Lipid droplet accumulation in differentiated cells was visualized using Oil Red O staining after 2.5 weeks of adipogenic induction. Calcium deposits were stained with Alizarin Red S after 2.5 weeks of osteogenic induction. The results show 1 representative sample from of 3 independent experiments.



Supplementary Figure S2. Hierarchical clustering analysis of normoxic- and hypoxic-cultured hUCB-MSCs.

A**B****C**

Supplementary Figure S3. Among the polycomb group proteins, BMI1 most effectively inhibits cellular senescence and improves the expression of COX-2. (A) After the overexpression of GFP, EZH2, SUZ12 and BMI1, hUCB-MSCs were subcultured and CPDLs were calculated. (B) After several passages, SA-β-gal staining was conducted to assess the senescent state of hUCB-MSCs overexpressing each factor. (C) GFP-, BMI1- and EZH2-transduced hUCB-MSCs were treated with IFN-γ and TNF-α for 24 hours, and western blot analyses were conducted to investigate the effects of the immunomodulatory proteins and COX-2 expression.

Table S1. Primers used in qRT-PCR and ChIP assay

qPCR Primers used for mRNA expression analysis

Target	Forward	Reverse
BMI1	GACTCTGGGAGTGACAAGGC	AGATTGGTGGTTACCGCTGG
p16INK4a	GAAGGTCCCTCAGACATCCC	CCCTGTAGGACCTTCGGTGA
DUSP1	CTGCCCTTTCTGTACCTGGG	GGTTGTCCTCCACAGGGATG
GAPDH	TGATGACATCAAGAAGGTGGTG	ACCCTGTTGCTGTAGCCAAAT

qPCR Primers used for ChIP analysis

Location	Forward	Reverse
p16INK4a promoter (-38 ~ -158)	GCACTCAAACACGCCTTTGC	AGAGCCAGCGTTGGCAAGGA
DUSP1 promoter 1 (-1815 ~ -1717)	GCGCCCAGCTCTTAAAAAGT	CCGACTTGATTTGTCCCATT
DUSP1 promoter 2 (-662 ~ -543)	GCTCGAGTCGGTCTTGGTAG	GACTTGCCCAGAACCACACT
DUSP1 promoter 3 (-106 ~ -16)	CCGTCACGTGATCACCATT	GCGTTTATATGCGGCCTCT